

A View-independent Line-Coding Colormap for Diffusion Tensor Imaging

Wu, Shin-Ting^a, Raphael Voltoline^a, Clarissa Lin Yasuda^b

^a*School of Electrical and Computer Engineering, University of Campinas, Campinas, Brazil*

^b*School of Medical Science, University of Campinas, Campinas, Brazil*

Abstract

Diffusion Tensor Imaging is a noninvasive technique promising for assessing the integrity of white matter tracts in the brain through the measurement of the movement of water. Because of the dimensions of the data involved, visualization of slice-by-slice images is still a challenge, and the colormaps for conveying the spatial direction of the major eigenvector of the diffusivity tensors widely adopted are ambiguous. The present paper addresses the issue of how to ameliorate this ambiguity. We propose a new line-coding color scheme, contemplating human visual perception in conjunction with the classic Hue-Saturation-Value color model. Experiments with neuroimages were also conducted to assess the potential of the proposal in the perception of spatial orientations in 2D views.

Keywords: Structural brain abnormalities visualization, Diffusion tensor imaging, Line-coding colormap

1. Introduction

The use of diffusion-weighted magnetic resonance images (DW-MRI) is a well-established technique for the estimation of the diffusivity of water in living tissues. This diffusion is anisotropic in organized tissues such as muscles and the white matter of the brain [1]. The introduction of diffusion tensor imaging (DTI) models has made it possible to describe this diffusivity in a way that is invariant, independent of rotation. Since its development, DTI has been applied in the investigation of various cerebral tract structures; this has increased the potential for the diagnosis of diverse neurological disorders [2]. The appropriate visualization of DTI images provides a potential technique for revealing the complex structure of the cerebral white matter in an understandable way [3].

In DTI the estimated diffusivity of a scanned sample is represented by a second-order tensor (3×3 matrix). Providing an integrated, but visually distinguishable, representation of all nine elements of each tensor is, however, not a trivial task. Kindlmann proposes superquadric functions for visualizing the tensors as spatial glyphs [4]. Since they can obscure each other, however, such glyphs fail to convey complex 3D fiber tracts. Fortunately, for most anatomical and functional studies, only the streamline of strongest diffusion is of interest, and the nine-element tensor visualization can be reduced to that of a single three-element vector. Such vectors can be used to construct 3D (hyper)streamlines with use of tractography [5, 6]. The result of this technique, however, is very sensitive to user input, and important information can be missed if the underlying anatomy is not known [7]. Furthermore, spatial paths are usually difficult to visualize from 2D images.

Slice-by-slice investigation is still the dominant practice among radiologists for the diagnosis of abnormalities [8]. In these 2D views of the slices, each sample can be viewed fully, thus allowing precise exploration and analysis. Multimodality

can enhance visualization for diagnosis, but this requires the unambiguous mapping of the fiber orientation onto some sort of graphics attribute to be displayed with other imaging modalities in order to improve the sensitivity and specificity of a 2D visual inspection protocol. Since 2D visual inspection continues to be used, the representation of both color and space in three dimensions can facilitate the portrayal of the vector fields as colors.

One of the uses of the measures of DTI is the identification of the paths of white matter tracts (line path), rather than a vector field of water diffusion (vector direction) [1]. Hence, in the visualization of DTI images, two co-linear vectors of opposite directions should be indistinguishable. Pajevic and Pierpaoli call such DTI data axial data, and they carefully compare the known Hue-Saturation-Value (*HSV*)-based color schemes in relation to the three main sources of misinterpretation: discontinuity artifacts, human perceptual distortions, and ambiguity in orientation. They conclude that none of the color schemes studied is free of misinterpretation. Figure 1 provides an example of the use of the *HSV* color scheme showing a unique representation, but suffering from discontinuity along the equator if the viewing angle is not restricted to less than 90° [9]. Figure 1(b) illustrates this discontinuity at the equator of a sphere with the points in the immediate vicinity mapped onto antipodal points. To illustrate this, Figure 1(b) presents these antipodal points in a rectangular coordinate system, with the vectors (x, y, z) presented as a solid line while $-(x, y, z)$ is dashed.

In this paper we present a novel color scheme which facilitates interpretation, while still perceptually satisfying the requirements of continuity and non-ambiguity. The key to this proposal is the exploration of smooth transitions between primary and secondary colors in the classic *RGB* color model. In Section 2 we compare the results of different color coding schemes in the visualization of DTI images. The proposed so-

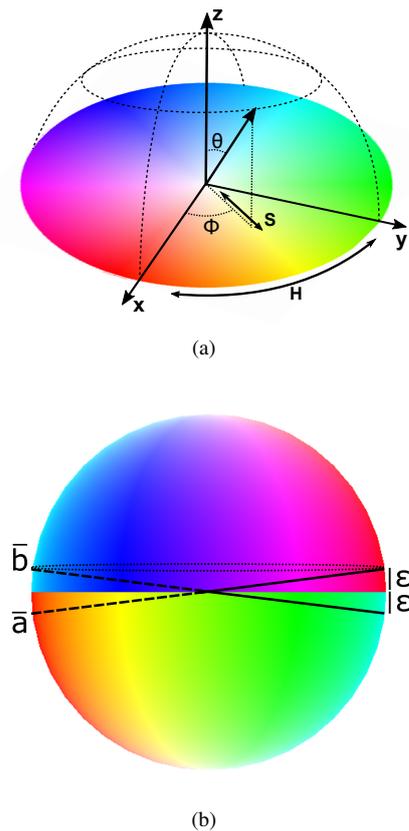


Figure 1: *HSV* mapping: (a) one-to-one mapping between color parameters (H, S) and the spherical coordinates (ϕ, θ) , and (b) discontinuity artifact on equator of the color sphere.

ors, with the difference between the colors becoming perceptually greater as the angle between the directions increases. The second requirement concerns the reference system of the color space, which should be invariant with respect to the angle of viewing, generally considered to be the orthogonal coordinate system of the patient. The third requirement refers to the common practice of modulating the color value by the anisotropy of diffusion, so that a sample becomes less visible as its isotropy decreases, a practice that can reduce visual pollution. The final one involves the mental inversion of color-mapped directions.

Each normalized 3D vector (x, y, z) in a patient's coordinate system must be unambiguously mappable to a distinct point on the unit sphere. Moreover, the vectors (x, y, z) and $-(x, y, z)$ must be mapped on the same color. Then, a single hemisphere of spherical coordinates (ϕ, θ) must be sufficient to represent all fiber orientations. Color coding of the spherical coordinates for such a point on a single hemisphere is possible if the *HSV* color model is used (Figure 1): the azimuthal angle ϕ and the polar angle θ of the direction of a fiber at each point are associated with a specific color hue (H) and the color saturation (S), respectively, through the following formula:

$$H = (\phi - \phi_R + 2\pi) \bmod 2\pi \quad S = \sin(\theta), \quad (1)$$

where $(\phi_R, \frac{\pi}{2})$ is the orientation of the spatial line that is mapped onto saturated red. For simplicity, the color value (V) is set at 1.0. Since ϕ_R is an arbitrary angle, modular arithmetic is adopted to avoid a negative H value: 2π is added to the difference $(\phi - \phi_R)$, and the result is wrapped around 2π . This mapping, denominated non-symmetrical color coding, is considered user-friendly, as it mimics the way an artist mixes paints on his palette [12]. Moreover, it does not suffer from ambiguity in orientation. However, it gives rise to color discontinuity artifacts, due to the mapping of two crossing lines, $\bar{a} = (\phi, \frac{\pi}{2} - \epsilon)$ and $\bar{b} = (\phi, \frac{\pi}{2} + \epsilon)$, on two distinct antipodal points in the *HSV* color space $((\phi, \frac{\pi}{2} - \epsilon)$ and $(\phi + \pi, \frac{\pi}{2} - \epsilon)$). In Figure 1(b) these two antipodal points lie on the dotted circle parallel to the equator. To overcome this discontinuity problem, preferred direction mapping is used.

Preferred direction mapping consists of applying non-symmetrical mapping on a unit sphere having its poles aligned in the preferred direction. Schlüter et al. propose the establishment of this direction in a way that automatically shifts the discontinuity artifacts. They define the preferred direction as the normal vector of the plane for which the average distance between the major diffusivity direction of each sample and its projection onto a plane is minimal [13]. This plane is called the optimal projection plane. Höller et al. map the angle between the average anisotropic diffusion direction and the vector perpendicular to the displayed slice onto the hue of the Hue-Saturation-Brightness (*HSB*) color model. The major shortcoming of the two preferred direction coding schemes, however, are that they violate the requirement of viewing invariance. Three other schemes for mapping have also been investigated in [9]: absolute value, rotational symmetry and mirror symmetry.

Absolute value mapping maps the absolute values of the ele-

lution is detailed in Section 3, and in Section 4 various experiments performed to assess the effectiveness of the proposed scheme in conveying information about the trajectory of neural tracts are outlined. The limited difference of this color mapping in relation to that observed in previous research makes this option seem promising, as discussed in Section 5. Finally, some concluding remarks are made in Section 6.

2. Related Work

Color-mapping is a useful and widely accepted technique for visualizing specific 3D vector fields, such as normal vectors [10] and the flow of fluids in computational fluid dynamics [11]. Since the seminal work of Pajevic and Pierpaoli [9], just a few suggestions of schemes for coloring DTI data using vector coding have been published. The main ideas presented have been summarized in this section.

According to Pajevic and Pierpaoli, four requirements are necessary in the design of a colormap for displaying fiber orientation: (1) linearity in color perception; (2) the reference system for describing orientation; (3) the establishment of a threshold for displayable anisotropy; and (4) the interpretability of the colors used. The first requirement is related to the use of color variation to indicate variations in vector direction. Vectors with similar directions should be portrayed by similar col-

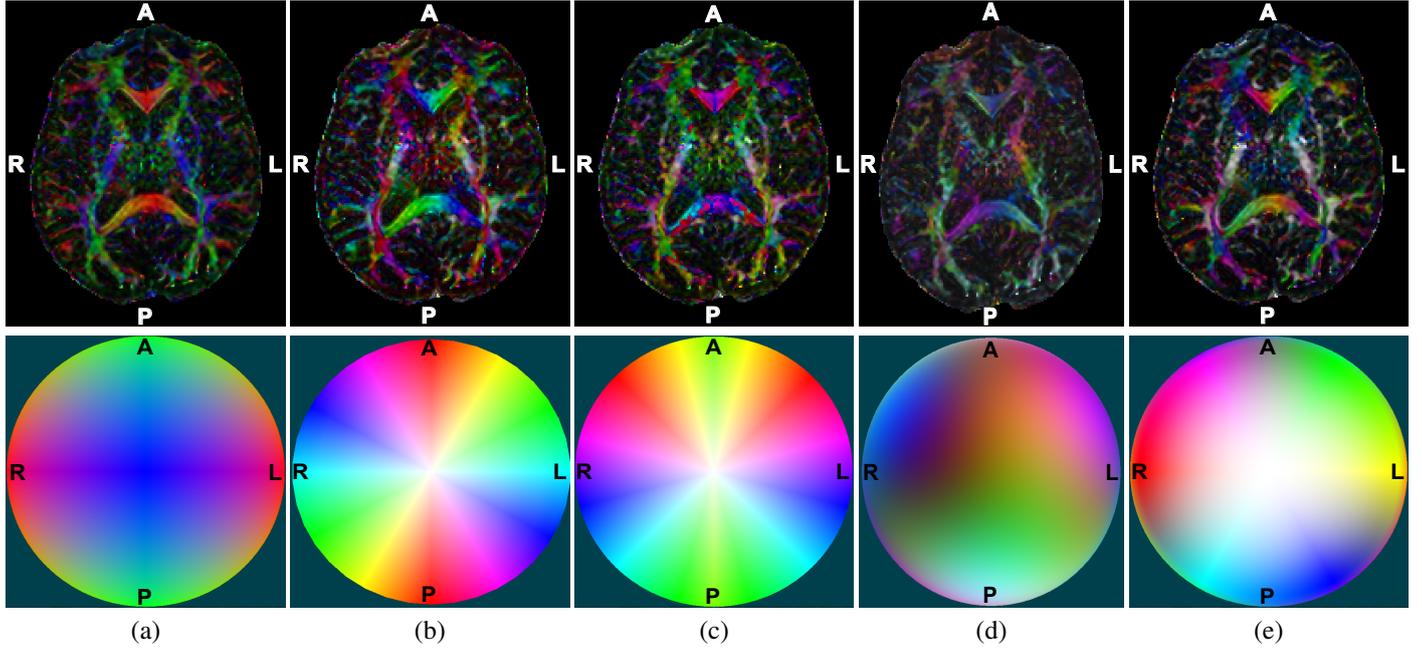


Figure 2: Different line-coding color schemes to represent the orientation of white matter fibers for each sample: (a) absolute value (conventional), (b) rotational symmetry, (c) mirror symmetry, (d) Boy surface immersion, and (e) the proposed color scheme. The DTI images showing color-coded orientation and the corresponding color coding schemes are shown in the top and the bottom row, respectively.

146 ments of fiber orientation (x, y, z) onto color elements, i.e.

$$R = |x| \quad G = |y| \quad B = |z|.$$

147 This color scheme has been found useful for displaying fiber
 148 tracts in axial slices, since it facilitates the discovery of the large
 149 asymmetries between the left and right hemispheres which provide
 150 evidence of abnormalities [14]. One example of this color
 151 coding is shown in the visualization of axial diffusion data in
 152 Figure 2(a). Note that there is no color discontinuity in regions
 153 where the variation in orientation is smooth, but there is a serious
 154 problem with ambiguity in spatial orientation. It is not possible,
 155 for example, to discern the difference in orientation of the
 156 commissural fibers in the left and right brain hemispheres.

157 Rotational symmetry mapping was designed to guarantee
 158 that any vector and its inverse are mapped onto the same color,
 159 i.e.

$$H = (2(\phi - \phi_R + 2\pi)) \bmod 2\pi.$$

160 Figure 2(b) shows how the ambiguity in orientation has been
 161 perceptually ameliorated, although not completely eliminated.

162 Mirror symmetry mapping was designed to portray any vector
 163 and its mirror image with respect to the midsagittal plane
 164 in the same color code, i.e. the azimuthal angle $\phi \in [0, \frac{\pi}{2}] \cup$
 165 $[\frac{3\pi}{2}, 2\pi]$ for each direction $(|x|, |y|, |z|)$ is mapped onto the specific
 166 hue using the following equation

$$H = 2((\phi - \phi_R + \pi) \bmod \pi).$$

167 Although mirror symmetry can be extremely helpful for high-
 168 lighting differences between the left and right brain hemisphere,

169 the mapping presents problems of ambiguity in orientation, as
 170 well as of discontinuity, as illustrated in Figure 2(c).

171 The color coding scheme proposed by He et al. was designed
 172 to maximize the use of the color space and reduce discontinuity
 173 artifacts [15]. These authors seem to resize and displace the di-
 174 rection vectors to the vectors in the first octant of the cartesian
 175 coordinate system, then to map the new vectors onto the RGB
 176 color cube; however, the fact that no key to the color mapping
 177 is provided in their paper and that the colors used are not famil-
 178 iar to neuroradiologists makes the interpretation of the images
 179 included in the paper quite difficult. It seems, however, that the
 180 proposed scheme satisfies the following unit vector (x, y, z) onto
 181 color (R, G, B) mapping rule:

$$R = \frac{x/2 + 0.5}{2} \quad G = \frac{y/2 + 0.5}{2} \quad B = \frac{z/2 + 0.5}{2}.$$

182 If this inference is correct, the scheme maps a vector (x, y, z)
 183 and its inverse $-(x, y, z)$ on two distinct colors, although this is
 184 not desirable for representing the pathways of fiber bundles.

185 The final color scheme presented is that proposed by Demiralp
 186 et al. for tackling the discontinuity problem on the equator
 187 of a sphere [16]. The key to their proposal is two-dimensional
 188 real projective space (RP^2) , which consists of the set of all lines
 189 in R^3 passing through the origin. This space is topologically
 190 equivalent to the unit sphere S^2 where every point and its anti-
 191 podal point are not distinguished, and can be further restricted
 192 to the upper hemisphere with the antipodal points at the equator
 193 “glued together”. There are some known immersions of RP^2 in
 194 R^3 : Boy surface, cross-cap and Roman surface.

Representing line paths in RP^2 , Demiralp et al. propose a
 “piecewise linear version of Boy’s surface” to map the line
 path in the rectangular coordinates (x, y, z) onto the coordinates

($f_1(x, y, z), f_2(x, y, z), f_3(x, y, z)$) of the *RGB* color space. This scheme first evaluates the function $g_j(x, y, z)$, $j = 1, 2, 3$, written as a partial weighted sum of $n + 1$ spherical harmonics

$$g_j(x, y, z) \approx \sum_{i=0}^n c_{ji} h_{ji}(x, y, z) \quad (2)$$

with $h_{ji}(x, y, z)$ and c_{ji} denoting the spherical harmonics and real-valued weights, respectively. Then, it scales and normalizes ($g_1(x, y, z), g_2(x, y, z), g_3(x, y, z)$) to get a point on the *RGB* space. It should be noted that some coefficients in Eq. 2 were adjusted by hand to achieve an aesthetically pleasing shape. An example implementation in Python is available online at [17].

The elegance of the underlying mathematical formulation of the color coding scheme proposed by Demiralp et al. and the generally unambiguously smooth mapping make the scheme extremely appealing, although there is a set of self-intersections due to R^3 realization. Nevertheless, inverse mapping from the colors onto the line paths constitutes a problem. As can be seen in Figure 3, it is not a trivial task to sequence the colors surrounding each reference axis to effectively convey variations in θ and ϕ in the Left-Posterior-Superior (LPS) patient reference system, i.e. *x*-axis for the patient’s L(ef) side, *y*-axis for the patient’s P(osterior) side and *z*-axis for the patient’s S(uperior) side [18]. Hence, fibers that have very similar spatial orientations may be colored with perceptually distinct colors, as is the case with the corticospinal tract in Figure 2(d). This does not comply with the requirement of linearity in color perception.

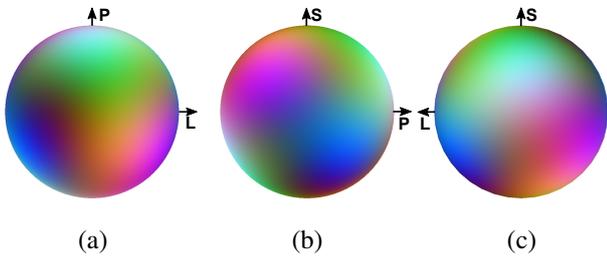


Figure 3: Orthogonal view of the Boy surface immersion color sphere in the direction of (a) S-axis, (b) L-axis and (c) P-axis.

Contributions The proposed alternative line-coding color scheme ameliorates ambiguity in orientation and visual discontinuity, thus facilitating the interpretation of DTI images, and it is useful in the more precise identification of the main direction of asymmetric water diffusion. Similar to the work of Demiralp et al. [16], this scheme presents a relatively unambiguous, smooth mapping between spatial orientations and colors. Instead of elegant but complex functions however, the present proposal relies on familiar color sequences and simple linear interpolation, thus facilitating the identification of spatial orientation from colors.

3. Proposal

Pajevic and Pierpaoli have noted that the main issue in unambiguous vector coding is the appearance of discontinuity [9].

Although the color wheel of the *HSV* color model can be easily interpreted and is familiar to most users, the discontinuity artifact due to discontinuity in the mapping in the vicinity of the equator can lead to misleading interpretations. Pajevic and Pierpaoli have also determined that perceptually uniform color spaces, within which the difference between the perception of two colors is proportional to their Euclidean distance, is not a solution, since that uniformity is limited when applied to color images. A huge number of color contrasts and discontinuity effects are known to influence visual experiences, but not because of perceptual non-uniformity. The illusion produced can even be an advantage and help reveal details and hidden information [19]. This background led us to look at the problem from a different angle, and the question was reformulated. Is it possible to remedy the flaws of *HSV*-based color codes without forfeiting their positive contributions?

The first idea was to change the position of the colors on the *HSV* color wheel so that the antipodal point of a primary color would become a secondary one, differing in respect to a single color element. This guarantees that a sequence of primary–secondary–primary–secondary colors is established with respect to the angle ϕ of line orientations and that the saturated hue transition is similar to that observed in the *HSV*-based color scheme.

However, it is clear in Figure 6(a) that the problem of the color discontinuity at the equator of the sphere remains. It thus seems reasonable to move the colors away from the equatorial borders of the hemispheres to a northern meridian and a southern meridian by λ degrees, with the interpolated colors of the hemisphere borders used to represent the lines that cross in the vicinity of the equator of the sphere to produce smooth perceptual transitions. Nevertheless, a little reflection shows that this approach will require certain additional modifications. Simple interpolation of arbitrary colors may result in many achromatic values, and a patient’s coordinate axis orientations may become perceptually indistinguishable.

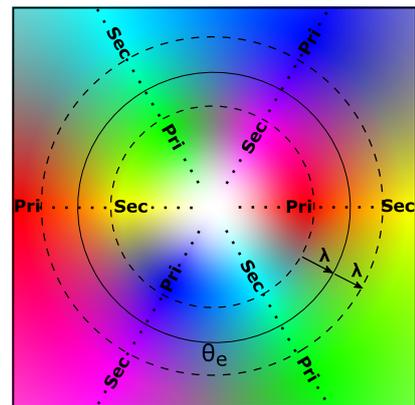


Figure 4: Proposed colormap without discontinuity artifact: arrangement of primary (Pri) and secondary (Sec) colors in stereographic projection.

It should be noted that in the present proposal the view-independent color codes for the patient’s reference axis orientations are no longer the usual red, green and blue hues presented in Figure 2(a). We show the new colors for the reference axes,

270 and suggest the adoption of a color weighting strategy to reduce
271 visual pollution.

272 3.1. Color Wheel

273 The proposal presented here involves a modification of the
274 spacing of colors around the color wheel. The primary and
275 secondary colors on the border of the color wheel are equally
276 spaced, counterclockwise, in the sequence red (1, 0, 0), magenta
277 (1, 0, 1), green (0, 1, 0), yellow (1, 1, 0), blue (0, 0, 1), and cyan
278 (0, 1, 1). This order is arbitrary, requiring only that, for each
279 saturated color, (a) one of its adjacent colors differs from it in a
280 single color element, while the other differs in three, and (b) the
281 color of the antipodal point differs by a single color element.

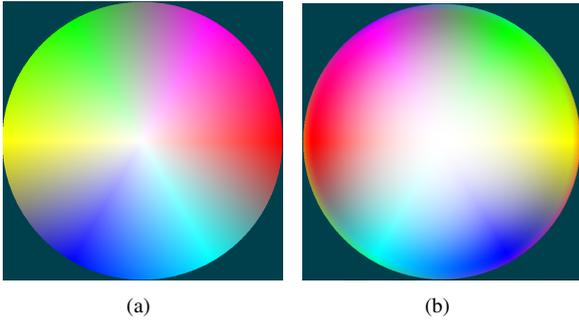


Figure 5: Combination of primary and secondary colors on the color wheel, in parallel projection, with different exponents of the saturation: (a) $n = 1$ and (b) $n = 2$.

282 Since the adjacent colors can differ in either one or three
283 color elements, only two rules for interpolation are necessary.
284 When two colors differ in respect to a single color element c ,
285 linear color variation is possible only from the color element
286 $c = 0.0$ to the color element $c = 1.0$. When they differ in three
287 elements (c_1, c_2, c_3), the pattern variation is: one color element
288 c_1 varies linearly from 0.0 to 1.0 and the other two elements
289 c_2 and c_3 also change linearly, but in the opposite direction,
290 from 1.0 to 0.0. From this interpolation, three samples along
291 the equator will be mapped onto light gray (0.5, 0.5, 0.5). How-
292 ever, color contrast effects will give the illusion of being differ-
293 ent colors, as will be discussed in Section 5.

Applying this colormap to the DTI images showed that, because of the contrast effect, the line parallel to the reference z-axis (coded in white) is almost indistinguishable in complex structures. This probably reflects the fact that our visual perception is one-third less sensitive to chromatic than to achromatic variations, and it is highly dependent on the surrounding colors [19]. To mitigate this perceptual limitation, the saturation of the color in the vicinity of the z-axis was reduced. Instead of Eq. 1, the color saturation is redefined as:

$$S = \sin(t^n * \theta_e), \quad (3)$$

where n is the exponent of the saturation and t , the saturation factor, given by

$$t = \begin{cases} \frac{\theta}{\theta_e - \lambda}, & \theta \leq (\theta_e - \lambda) \\ 1, & \text{elsewhere} \end{cases},$$

294 where θ_e and λ are, respectively, the polar coordinate of the
295 equator of the sphere and the interpolation range, as will be
296 discussed in Section 3.2.

297 Figure 5 shows the difference in the colormaps generated
298 with the exponent of saturation $n = 1$ (a) and with $n = 2$ (b). In
299 this paper, all DTI slices are rendered with $n = 2$.

300 3.2. Linear Interpolation

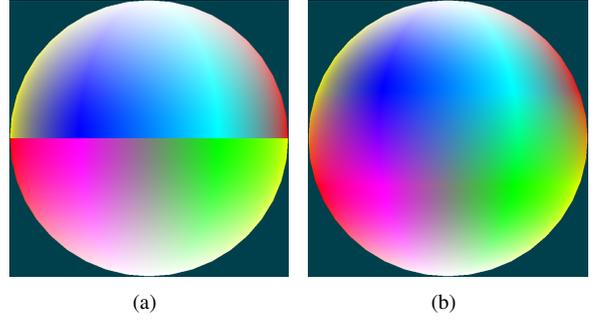


Figure 6: Interface of two orientation hemispheres: (a) without and (b) with the interpolation belt on the equator.

301 With respect to the sphere equator, depicted as a solid circle
302 in Figure 4, the adjacent colors located in the two different
303 hemispheres differ in a single color element. Even so, how-
304 ever, the transition between them is not perceptually smooth, as
305 shown in Figure 6(a). To remedy this problem, a transition belt
306 on the two sides of the equator is defined between $\theta_e - \lambda$ and
307 $\theta_e + \lambda$, with θ_e being the polar coordinate of the equator. In Fig-
308 ure 4 this region is bounded by the dashed circles. The colors
309 on the displaced border of the northern hemisphere and those on
310 the displaced border of the southern hemisphere are located in
311 the inner and the outer dashed circles, respectively. The colors
312 in the belt bounded by these two dashed circles are interpolated
313 in a way analogously to what was done with the adjacent col-
314 ors on the color wheel: if the values of the color elements of
315 two colors are different, the color between them is subjected to
316 linear interpolation, thus resulting in a smooth transition. Fig-
317 ure 6(b) shows the result of interpolation for $\lambda = 20^\circ$.

318 3.3. Axis Orientation

319 The color code for the orientation of the reference axis is
320 crucial in the interpretation of color. To be view-independent,
321 the reference system is generally aligned with the LPS-patient
322 system. As already explained in Section 3.1, the line parallel
323 to the z-axis, the S-axis, is coded in white (Figure 7(a)). Both
324 the line parallel to the L-axis and that parallel to the P-axis are
325 color-coded along the equator of the color sphere (Section 3.2).
326 In the proposed scheme, the L-axis is represented by a shade
327 of orange (the interpolation of red and yellow, corresponding
328 to the displaced border colors at $\phi = 0$), and the P-axis by a
329 shade of blue (the interpolation of the displaced border colors
330 at $\phi = 90^\circ$). In Figure 7(b) the color sphere is rotated so that
331 the point corresponding to the L-axis is located in the center of
332 the color disc, while in Figure 7(c) this central point represents
333 the P-axis.

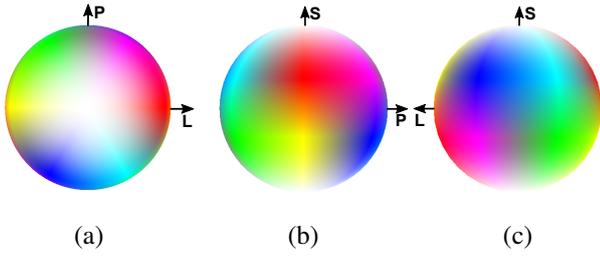


Figure 7: Orthogonal view of the color sphere in the direction of (a) S-axis, (b) L-axis and (c) P-axis.

3.4. Encoding Anisotropy of Diffusion

As mentioned in Section 2, it is common to weight the color vector by a scalar value characterizing diffusion. This scalar value can be the fractional anisotropy (FA) [20], which lies in the interval $[0.0, 1.0]$. When using FA to weight a color, the lower the FA, the darker the sample color is, with a decrease in FA from 1.0 to 0.0 resulting in a diminished hue. Therefore, nearly isotropic samples are either dark gray or black in color, and they are perceptually filtered out against a black background. This is the simplest way to highlight neural fibers. This filtering technique has been applied for all the DTI slices presented in this paper.

4. Experiments

The proposed color coding scheme was evaluated with diffusion data in relation to effectiveness in conveying the major neural tracts and its potential clinical value. This section presents the results from the data volumes of a healthy volunteer, as well as those of patients with neural disorders. The DTI images used here were obtained from DW-MRI images with 70 axial slices of 256×256 . They were acquired with a Philips Achieva 3T Scanner using a spin-echo echo-planar diffusion-weighted sequence, 32 diffusion-encoding gradients, and a b-value of 1000 s/mm^2 . The DW-MRI data acquired were processed using the FSL Diffusion Toolkit [21]. Because the raw data are in the format of Digital Imaging and Communications in Medicine (DICOM) [22], they were converted to the NIfTI format with dcm2nii software [23]. It should be noted that all subjects enrolled in the present study signed the informed consent form approved by the Ethics Committee of our university.

To assess the power of the proposed color scheme for clarity in the visualization of the line path, a study was made of certain tracts that are frequently studied with DTI images to compare those created applying the most frequently used absolute values with those based on the proposed color scheme (Figures 8–10). The original study of Pajevic and Pierpaoli had already compared the results of absolute values to those of other color schemes [9]. In Figures 8–10 the slices in the top row are colored with the traditional absolute value color scheme, whereas in the bottom row the same slices are rendered using the scheme proposed here.

Figure 8 shows the colormap of the major commissural fiber system, the corpus callosum, which connects the two cerebral

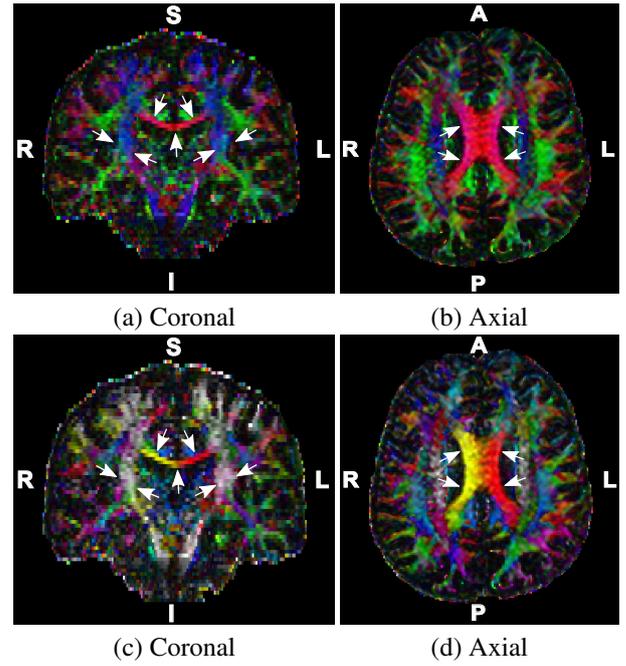


Figure 8: Comparison of fiber tracts in the corpus callosum, colored using absolute value (top row) and the proposed scheme (bottom row).

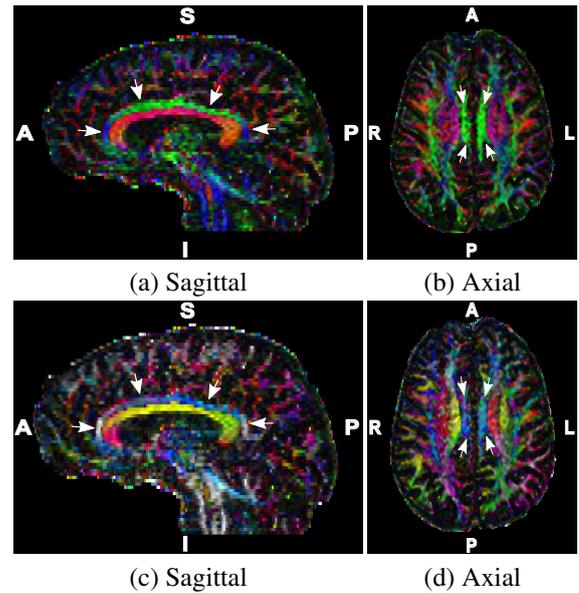


Figure 9: Comparison of fiber tracts in the cingulum, colored using absolute value (top row) and the proposed scheme (bottom row).

hemispheres in a right-to-left orientation, as well as the corticospinal tracts. The leftmost column and the rightmost one illustrate, respectively, coronal slices with the corticospinal tracts and axial slices with the maximum anterior-to-posterior width of the corpus callosum (indicated by the white arrow). Despite the low acquisition resolution along the axis from feet to head, which made angular variations difficult to distinguish in the coronal slice, the corticospinal tracts, in the shade of blue in the absolute value color scheme (Figure 8(a)), are more clearly revealed by the whitish color in Figure 8(c) with the present

386 scheme. Moreover, the extremes of the corpus callosum bend
 387 up toward the patient's superior side, but the orientation of the
 388 two is different. This can be seen in the transitional orange of
 389 the present scheme (Figures 8(d)) located between the red indi-
 390 cating fibers running to the upper left and the yellow for those
 391 running to the upper right, in contrast to the single shade of red
 392 in the absolute color scheme (Figure 8(b)). It is remarkable that
 393 the ambiguities in orientation revealed in Figure 8(b) have been
 394 eliminated, as indicated by the white arrows in Figure 8(d).

395 Figure 9 shows the colormap of the cingulum (indicated by
 396 the white arrows). The left and the right columns present,
 397 respectively, sagittal and axial slices, showing the maximum
 398 anterior-to-posterior width of the cingulum. The cingulum is
 399 a collection of white matter fibers projecting from the cingu-
 400 late gyrus to the entorhinal cortex. It is C-shaped, wrapping
 401 the corpus callosum from the frontal lobe to the temporal lobe.
 402 The continuous variations in colors are difficult to perceive in
 403 the absolute value scheme (Figures 9(a) and (b)). But, in the
 404 proposed scheme, we can see from Figures 9(c) and (d) that the
 405 fibers run in an anterior-to-posterior direction, with the inferior-
 406 to-superior directions varying gradually (from a superior orien-
 407 tation (white), to posterior-and-superior (grayish magenta) and
 408 then to anterior-and-inferior (light blue)). These colors can be
 409 seen to vary smoothly along the path from white to light blue.
 410 This produces the perception that the tract curves continuously
 411 along the path, even given the low resolution of the sagittal
 412 slice.

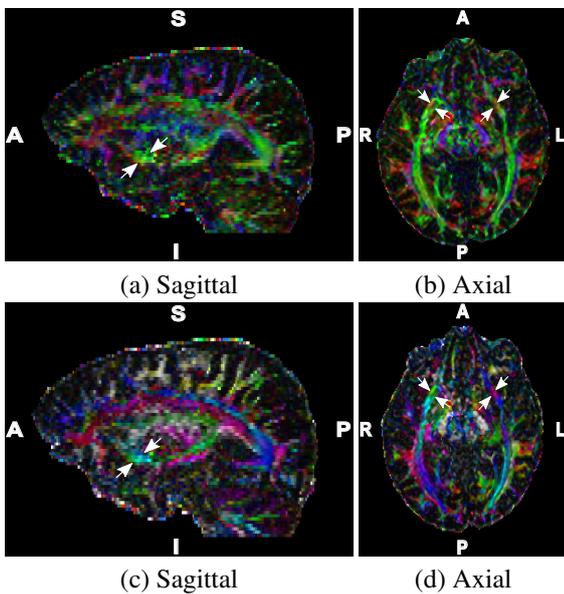


Figure 10: Comparison of fiber tracts in the uncinate fasciculus, colored using absolute values (top row) and those of the proposed scheme (bottom row).

413 Figure 10 shows the uncinate fasciculus, which is a white
 414 matter tract that connects the hippocampus and amygdala in
 415 the temporal lobe to the frontal lobe. It is a hook-shaped bun-
 416 dle running in an anterior-to-posterior orientation. Sagittal and
 417 axial slices colored with the two color schemes are portrayed
 418 in the left- and right-hand columns of the figure. In the two
 419 color schemes, the tracts are barely visible. In the slices col-

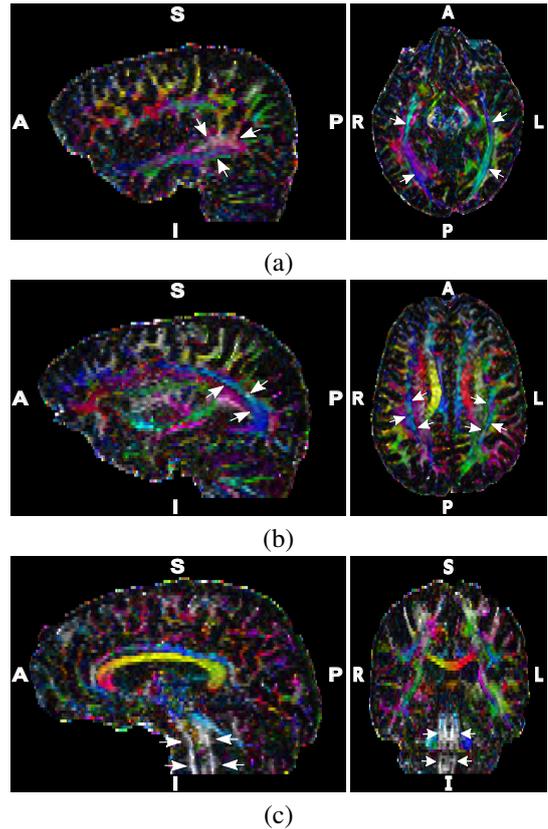


Figure 11: Three major neural tracts: (a) inferior, (b) superior longitudinal fasciculus; and (c) corticospinal tract.

420 ored using the absolute values (Figures 10(a) and (b)), this tract
 421 is displayed with a shade of greenish orange, while with the
 422 proposed scheme (Figures 10(c) and (d)), the origin lateral to
 423 the amygdala and hippocampus in the temporal lobe is seen to
 424 curve upward behind the external capsule and inward along the
 425 insular cortex (bluish shade), then up into the posterior part of
 426 the orbital gyrus (a shade of magenta). The white arrows in
 427 Figure 10 indicate this anatomic structure.

428 The next visualization shows the orientation of three major
 429 neural tracts with the proposed color scheme: the inferior and
 430 the superior longitudinal fasciculi and the corticospinal tracts.
 431 The inferior longitudinal fasciculus consists of a pair of tracts
 432 that run along the lateral ventricle, connecting the occipital and
 433 the temporal lobes, while the superior longitudinal fasciculus
 434 connects the front and back of the cerebrum. The corticospinal
 435 tract runs longitudinally. The white arrows in Figures 11 high-
 436 light the structures of interest. The tracts of the inferior longi-
 437 tudinal fasciculus reveal mirror symmetry, but not exactly paral-
 438 lel; the left bundle is coded in a shade of bluish green while the
 439 right-hand bundle is in cyanish magenta (Figure 11(a)). Since
 440 the bundles of the superior longitudinal fasciculus are almost
 441 parallel to the cingulum along its entire course; they are coded
 442 throughout in the same bluish shade as the cingulum. This can
 443 be seen in Figure 11(b). In Figure 11(c) the corticospinal tract
 444 is colored white, corresponding to the orientation of the spinal
 445 cord from feet to head.

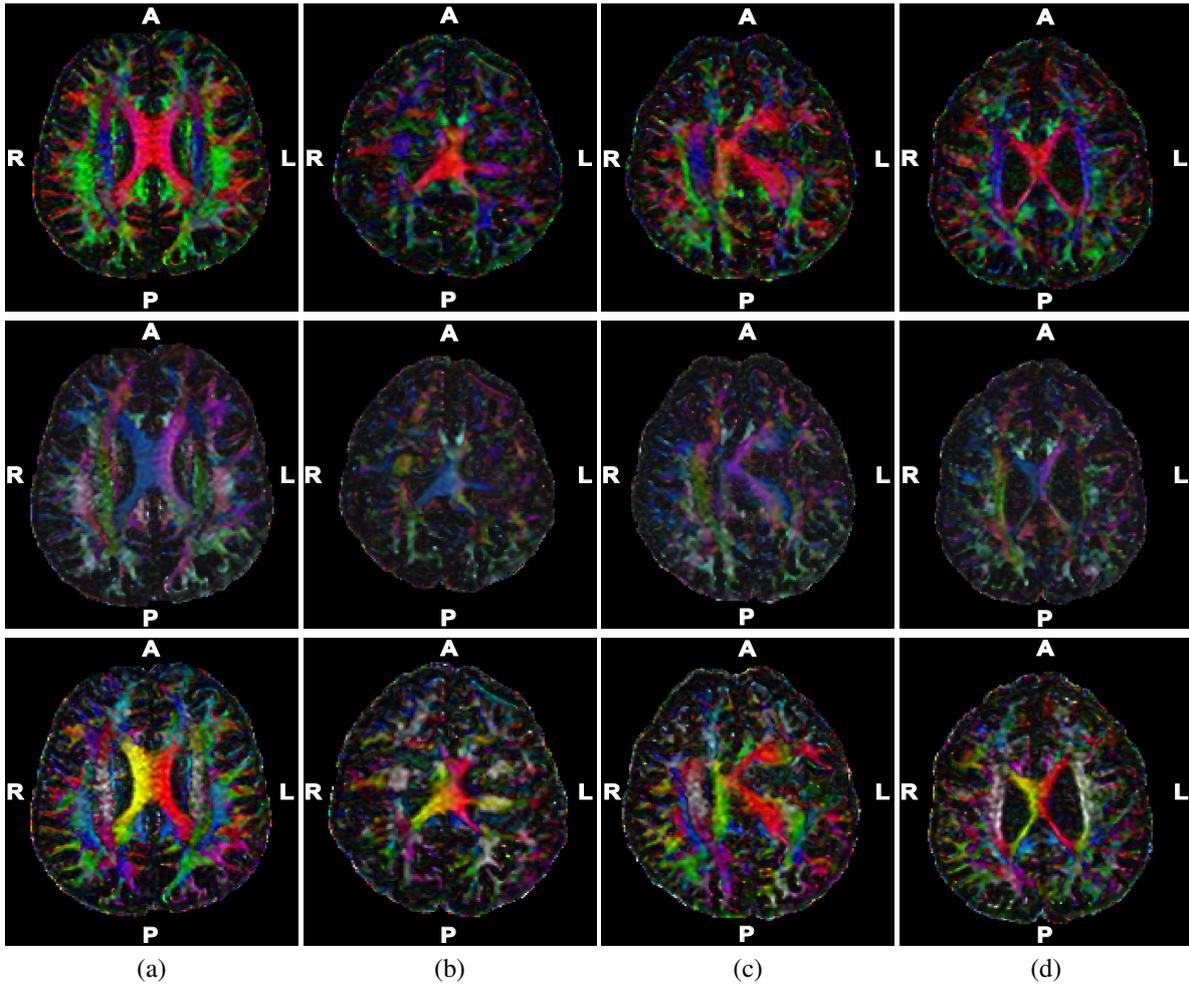


Figure 12: Axial slices at the point of maximum anterior-to-posterior width of the corpus callosum: absolute value color scheme in the top row, Boy surface scheme in the middle row, and the proposed color scheme in the bottom row. As a reference, (a) shows a healthy control. The other columns (b)–(d) display slices from patients with brain malformations. The color patterns in the bottom row draw more attention to structural abnormalities.

446 Figure 12 provides a comparison of the results of the abso-
 447 lute value scheme (top row), those of the Boy surface scheme
 448 (middle row), and those of the proposed color scheme (bottom
 449 row) in the revelation of spatial orientation of the tract in the
 450 DTI exams of patients with subcortical band heterotopia. This
 451 congenital disease consists of a diffuse neuronal migration that
 452 leads to the anomalous presence of gray matter interspersed in
 453 the white matter. For comparison, the colored slices of a healthy
 454 control are shown in Figure 12(a). The proposed color scheme
 455 (bottom row) reveals more details than do the absolute value
 456 and Boy surface scheme, not only for the healthy control, but
 457 also when abnormalities are present. The abnormality is espe-
 458 cially clear in the slice of the corpus callosum colored using
 459 the proposed scheme, where the fibers which should be running
 460 upwards (red and yellow in the normal brain) are revealed to be
 461 running in other directions (greenish and magenta). The spatial
 462 orientation of the superior longitudinal fasciculus is also more
 463 discernible with the proposed scheme than it is with the Boy
 464 surface scheme.

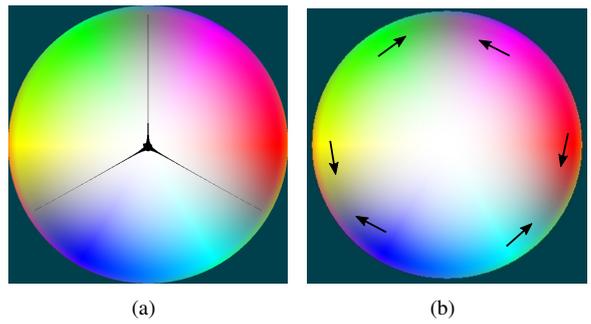


Figure 13: Pitfalls of the proposed color scheme: (a) non-invertibility and (b) perceptually gray bands.

465 5. Discussion

466 This paper addresses the issue of the ambiguity found in the
 467 color coding of spatial line path and proposes a less ambiguous
 468 option. In order to assign distinctive and smoothly varying col-
 469 ors to line orientations, we propose a color wheel consisting of
 470 alternating primary and secondary colors as illustrated in Fig-

471 ure 4. When compared with previous research, the proposed
 472 color scheme furnishes a representation with a perceptually un-
 473 ambiguous line orientation without the artifact of discontinu-
 474 ity. Nevertheless, similar to the results with the Boy surface
 475 scheme, this mapping is not invertible for all colors. This is
 476 because in the interpolation of a primary color and a secondary
 477 color differing in three elements (such as red (1.0, 0.0, 0.0) and
 478 cyan (0.0, 1.0, 1.0)), the same gray values are obtained at three
 479 points along each latitude of the color sphere. All the orienta-
 480 tions highlighted in black in Figure 13(a) are ambiguously
 481 mapped. Therefore, the proposed color scheme is not appropri-
 482 ate for interactions such as color picking for the identification
 483 of a specific orientation, although an evaluation of the colors
 484 in the vicinity of the ambiguous gray does provide a possible
 485 solution for distinguishing the coded orientations.

486 Although the linear interpolation of adjacent colors leads to
 487 the three gray bands on the color sphere indicated by the black
 488 arrows in Figure 13(b) can be argued to cause incorrect inter-
 489 pretation of line path, the consideration of these colors in con-
 490 junction with the adjacent ones leads to a different perception
 491 of the DTI images. Our explanation for these surprising results
 492 is that the appearance of a color is strongly dependent on its
 493 surroundings [19] and the smoothness of the geometry of tract.

494 To validate the proposed line-coding color scheme, both co-
 495 registered DTI and T1-weighted MRI images were used to as-
 496 certain the location of the uncinate fasciculus as it runs upwards
 497 from the temporal lobe to the insular cortex to form a spatial
 498 pathway that intersects the 2D slices under investigation diagon-
 499 ally, rather than transversely or longitudinally. Therefore, the
 500 many intersections are reduced to a set of points on the slices,
 501 which makes it difficult to identify the uncinate fasciculus in a
 502 slice-by-slice investigation. A 3D visualization, however, can
 503 help clarify the spatial orientation of these white matter tracts.

504 Despite the shortcomings in the color codes mentioned a-
 505 bove, it is still widely used in data visualization, either because
 506 it approximates the physical spectrum of applications or has a
 507 direct correspondence to the vectors represented. The famili-
 508 arity of 2D visualization makes combination with colored DTI
 509 volumes an interesting option for multimodal visualization. Far
 510 from providing an accurate visualization of spatial orientation,
 511 the proposed color scheme does provide a simple and rapid al-
 512 ternative furnishing more information about the orientation of
 513 fiber tracts than do the traditional color schemes.

514 The way to display the key for the color mapping of the
 515 path of a spatial line on a plane presents a challenge. This is-
 516 sue has also been addressed. Three colored sphere projections
 517 were investigated: stereographic projections (Figure 4), paral-
 518 lel ones (Figure 5(b)), and equirectangular ones (Figure 14).
 519 From our corridor testing, all three colormaps required extra
 520 mental effort for interpretation. Nevertheless, possibly because
 521 of our pre-attentive association of a disc-shaped figure with a
 522 sphere, it seems that the disc shape produces better spatial per-
 523 ception. The stereographic and parallel projections were found
 524 to be perceptually similar. Although equirectangular projection
 525 is not suitable as the key to color mapping, it is quite appropri-
 526 ate for texture lookup-based implementation. A comparison of
 527 the equirectangular projection of the proposed scheme in Fig-

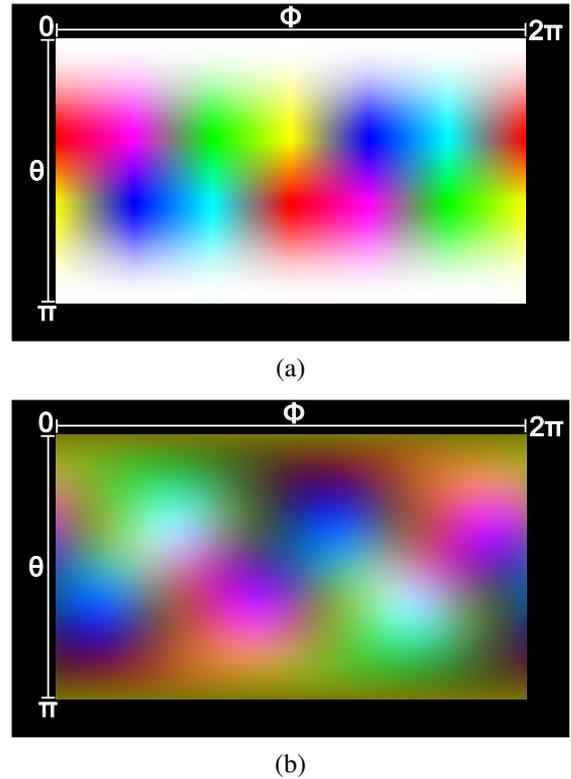


Figure 14: Equirectangular projection as the key to color mapping of line paths: (a) present proposal; (b) Boy surface scheme (available at [24]).

528 ure 14(a) and that provided by the Boy surface scheme (Fig-
 529 ure 14(b)) suggests that the proposed scheme is preferable in
 530 relation to color sequencing and richness of tone.

531 6. Concluding Remarks

532 Although the 3D rendering of fiber tracts has evolved rapidly,
 533 scalar indices such as fractional anisotropy (FA) and mean dif-
 534 fusivity (MD) are still the most widely used tools in the study
 535 of the microstructural properties of tissues. 2D visualization is
 536 familiar to radiologists, even though the path of fibers cannot
 537 be conveyed. Such information about orientation can be cru-
 538 cial for understanding the architecture of fibers, however, and
 539 a novel option has been proposed here, one based on a line-
 540 coding color scheme for displaying a fiber tract in 2D. A cor-
 541 ridor test was performed to obtain the feedback of both physi-
 542 cians and non-physicians. All volunteers could distinguish cor-
 543 rectly the direction of major diffusivity once the color display
 544 had been explained. Such a superficial approach, however, can-
 545 not demonstrate the true clinical value of the scheme. In the
 546 future, we plan to design an usability test for potential users
 547 who are familiar with DTI images to assess the utility of the
 548 proposed scheme.

549 As already discussed in Section 5, the proposed color coding
 550 scheme is not adequate for all studies of tract architecture, in-
 551 cluding the analysis of the symmetry of cerebral hemispheres.
 552 The proposed scheme is, however, a complementary scheme

553 designed to facilitate the study of subtle variations in diffusiv-
 554 ity in slice-by-slice exploration. We believe that, together with
 555 other vector color schemes and tractography, one can build a
 556 3D exploratory visualization environment that supports the Vi-
 557 sual Information Seeking Mantra [25]: (3D) Overview first,
 558 zoom and filter, then (2D) details-on-demand. Such a visual
 559 exploratory environment is our long-term goal.

560 Acknowledgements

561 The authors would like to acknowledge the contribution of
 562 Fernando Cendes in providing the MRI volume data and of
 563 Linda G. El-Dash in the proof-reading of the text. The research
 564 was supported by a CNPq-Brazil fellowship (305785/2012-5,
 565 308764/2015-3), a CNPq-Brazil scholarship (153389/2014-1),
 566 and the Fapesp-Brazil grant #2013/07559-3 to the BRAINN
 567 Research, Innovation and Dissemination Center of the Univer-
 568 sity of Campinas.

569 References

570 [1] Mori S, van Zijl PCM. Fiber tracking: principles and strategies - a
 571 technical review. *NMR in Biomedicine* 2002;15(7-8):468–80. URL:
 572 <http://dx.doi.org/10.1002/nbm.781>. doi:10.1002/nbm.781.

573 [2] Lerner A, Mogensen MA, Kim PE, Shiroishi MS, Hwang
 574 DH, Law M. Clinical applications of diffusion tensor imag-
 575 ing. *World Neurosurgery* 2014;82(1-2):96–109. URL:
 576 <http://dx.doi.org/10.1016/j.wneu.2013.07.083>.
 577 doi:10.1016/j.wneu.2013.07.083.

578 [3] Mori S, Zhang J. Principles of diffusion tensor imaging and its ap-
 579 plications to basic neuroscience research. *Neuron* 2006;51(5):527–
 580 39. URL: <http://dx.doi.org/10.1016/j.neuron.2006.08.012>.
 581 doi:10.1016/j.neuron.2006.08.012.

582 [4] Kindlmann G. Superquadric tensor glyphs. In: Deussen O, Hansen
 583 C, Keim D, Saupé D, editors. *Eurographics / IEEE VGTC Symposi-
 584 um on Visualization*. Aire-la-Ville, Switzerland, Switzerland: The
 585 Eurographics Association. ISBN 3-905673-07-X; 2004, p. 147–54.
 586 URL: <http://dx.doi.org/10.2312/VisSym/VisSym04/147-154>.
 587 doi:10.2312/VisSym/VisSym04/147-154.

588 [5] Bihan DL, Mangin JF, Poupon C, Clark CA, Pappata S, Molko
 589 N, et al. Diffusion tensor imaging: Concepts and applica-
 590 tions. *J Magn Reson Imaging* 2001;13(4):534–46. URL:
 591 <http://dx.doi.org/10.1002/jmri.1076>. doi:10.1002/jmri.1076.

592 [6] Jones DK, Travis AR, Eden G, Pierpaoli C, Basser PJ. PASTA:
 593 Pointwise assessment of streamline tractography attributes.
 594 *Magnetic Resonance in Medicine* 2005;53(6):1462–7. URL:
 595 <http://dx.doi.org/10.1002/mrm.20484>. doi:10.1002/mrm.20484.

596 [7] Vilanova A, Zhang S, Kindlmann G, Laidlaw D. An introduction to vi-
 597 sualization of diffusion tensor imaging and its applications. In: *Visual-
 598 ization and Image Processing of Tensor Fields*. Springer-Verlag; 2006,
 599 p. 121–53. URL: <http://dx.doi.org/10.1007/3-540-31272-2.7>.
 600 doi:10.1007/3-540-31272-2.7.

601 [8] Preim B, Botha CP. *Visual Computing for Medicine: Theory, Algorithms,
 602 and Applications*. 2 ed.; San Francisco, CA, USA: Morgan Kaufmann
 603 Publishers Inc.; 2013. ISBN 9780124159792.

604 [9] Pajevic S, Pierpaoli C. Color schemes to represent the orienta-
 605 tion of anisotropic tissues from diffusion tensor data: applica-
 606 tion to white matter fiber tract mapping in the human brain.
 607 *Magnetic resonance in medicine* 1999;42(3):526–40. URL:
 608 [http://dx.doi.org/10.1002/\(SICI\)1522-2594\(199909\)-
 609 42:3<526::AID-MRM15>3.0.CO;2-J](http://dx.doi.org/10.1002/(SICI)1522-2594(199909)42:3<526::AID-MRM15>3.0.CO;2-J). doi:10.1002/(SICI)1522-
 610 2594(199909)42:3<526::AID-MRM15>3.0.CO;2-J.

611 [10] Engel K, Hadwiger M, Kniss J, Rezk-Salama C, Weiskopf D. *Real-
 612 time Volume Graphics*. Natick, MA, USA: A. K. Peters, Ltd.; 2006.
 613 ISBN 1568812663. URL: <http://dx.doi.org/10.1201/b10629>.
 614 doi:10.1201/b10629.

615 [11] Post FH, van Walsum T. Fluid flow visualization. In: *Fo-
 616 cus on Scientific Visualization*. London, UK, UK: Springer-
 617 Verlag. ISBN 3-540-54940-4; 1993, p. 1–40. URL:
 618 http://dx.doi.org/10.1007/978-3-642-77165-1_1.
 619 doi:10.1007/978-3-642-77165-1_1.

620 [12] Smith AR. Color gamut transform pairs. *ACM
 621 SIGGRAPH Computer Graphics* 1978;12(3):12–9.
 622 URL: <http://dx.doi.org/10.1145/965139.807361>.
 623 doi:10.1145/965139.807361.

624 [13] Schlüter M, Stieltjes B, Rexilius J, Hahn H, Peitgen HO. Unique
 625 planar color coding of fiber bundles and its application to fiber in-
 626 tegrity quantification. In: *2004 2nd IEEE International Symposium
 627 on Biomedical Imaging: Macro to Nano (IEEE Cat No. 04EX821);
 628 vol. 1*. Institute of Electrical & Electronics Engineers (IEEE); 2004, p.
 629 900–3. URL: <http://dx.doi.org/10.1109/ISBI.2004.1398684>.
 630 doi:10.1109/isbi.2004.1398684.

631 [14] Liu SX. Symmetry and asymmetry analysis and its impli-
 632 cations to computer-aided diagnosis: A review of the litera-
 633 ture. *Journal of Biomedical Informatics* 2009;42(6):1056–64.
 634 URL: <http://dx.doi.org/10.1016/j.jbi.2009.07.003>.
 635 doi:10.1016/j.jbi.2009.07.003.

636 [15] He R, Mehta M, Narayana P. Color coding for visualization of the di-
 637 rectional information of DTI. In: *The 26th Annual International Confer-
 638 ence of the IEEE Engineering in Medicine and Biology Society; vol. 1*.
 639 Institute of Electrical & Electronics Engineers (IEEE); 2004, p. 1857–
 640 9. URL: <http://dx.doi.org/10.1109/IEMBS.2004.1403552>.
 641 doi:10.1109/iembs.2004.1403552.

642 [16] Demiralp C, Hughes J, Laidlaw D. Coloring 3d line
 643 fields using boy’s real projective plane immersion. *IEEE
 644 Trans Visual Comput Graphics* 2009;15(6):1457–64.
 645 URL: <http://dx.doi.org/10.1109/tvcg.2009.125>.
 646 doi:10.1109/tvcg.2009.125.

647 [17] Opensource . Diffusion mr imaging in python. 2016. URL:
 648 [https://github.com/nipy/dipy/blob/master/dipy/viz/co-
 649 lormap.py](https://github.com/nipy/dipy/blob/master/dipy/viz/colormap.py); accessed in July 2016.

650 [18] Wideman G. Orientation and Voxel-Order Termi-
 651 nology: RAS, LAS, LPI, RPI, XYZ and All That.
 652 [http://www.grahamwideman.com/gw/brain/orientation/o-
 653 rientterms.htm](http://www.grahamwideman.com/gw/brain/orientation/orientterms.htm); 2016. Accessed in February 2016.

654 [19] Ware C. Color sequences for univariate maps: theory, experiments
 655 and principles. *IEEE Comput Grap Appl* 1988;8(5):41–9. URL:
 656 <http://dx.doi.org/10.1109/38.7760>. doi:10.1109/38.7760.

657 [20] Pierpaoli C, Basser PJ. Toward a quantitative assessment of diffusion
 658 anisotropy. *Magn Reson Med* 1996;36(6):893–906.

659 [21] FMRIB . FMRIB’s Diffusion Toolbox. 2016. URL:
 660 <http://fsl.fmrib.ox.ac.uk/fsl/fslwiki/FDT>; accessed in
 661 February 2016.

662 [22] NEMA . DICOM – Digital Imaging and Communications in Medicine.
 663 2016. URL: <http://dicom.nema.org/>; accessed in February 2016.

664 [23] DCM2NII . dcm2nii DICOM to NIfTI conversion. 2016. URL:
 665 [http://www.mccauslandcenter.sc.edu/micro/mricron/dcm-
 666 2nii.html](http://www.mccauslandcenter.sc.edu/micro/mricron/dcm2nii.html); accessed in February 2016.

667 [24] Voltoline R. Multimodal visualization of diffusion tensor imaging. 2016.
 668 URL: [http://www.dca.fee.unicamp.br/projects/mtk/volto-
 670 line/download.html](http://www.dca.fee.unicamp.br/projects/mtk/volto-

 669 line/download.html); accessed in July 2016.

671 [25] Shneiderman B. The eyes have it: a task by data type taxonomy for
 672 information visualizations. In: *Proceedings 1996 IEEE Symposium on
 673 Visual Languages. VL ’96*; Washington, DC, USA: Institute of Elec-
 674 trical & Electronics Engineers (IEEE). ISBN 0-8186-7508-X; 1996,
 675 p. 336–43. URL: <http://dx.doi.org/10.1109/VL.1996.545307>.
 676 doi:10.1109/vl.1996.545307.